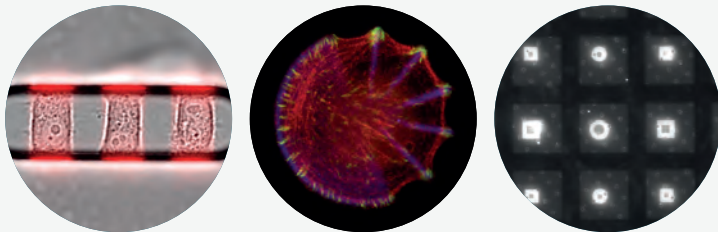




Better cell sample preparation
for cell biology & cryo-ET

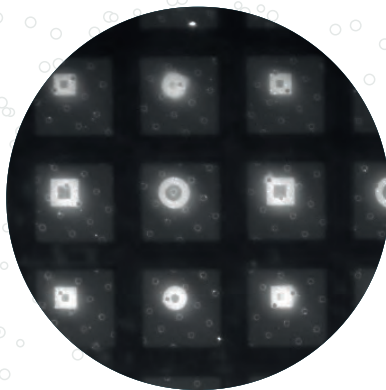


alvéole 

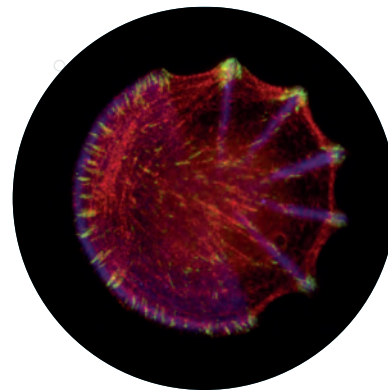
Better cell sample preparation, Better *in vitro* cell models.

One of the challenges confronting cell biologists *in vitro* is to work with **controlled and reproducible microenvironments** to more efficiently study living cells and model diseases.

PRIMO2 contactless and maskless photopatterning platform allows to engineer custom *in vitro* microenvironments and **fine-tune their mechanical and biochemical properties**, at the micrometer scale with high flexibility and reproducibility.



ECM protein micropatterns on holey carbon EM grids (200 mesh gold grid bars) done with PRIMO maskless photopatterning for controlling the cell adhesion location. L. Engel et al., JMM, 2019



Embryonic fibroblasts from vimentin knockout mice on fibronectin + fibrinogen-A647 (blue) micropattern, actin (red) and focal adhesions (green). Courtesy of A.J. Jimenez and B. Vianay, Physics of cytoskeleton & Morphogenesis lab

Benefits



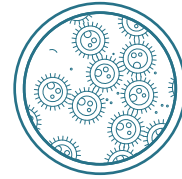
Time saving & independence

Rapidly optimize your experimental conditions yourself.



High Flexibility

Download any images you want to structure and/or functionalize your substrate.



Versatility

Use your regular cell culture substrates: stiff or soft, flat or microstructured.



My interest is to understand the role of biophysical and topological properties of tissue microenvironments, such as stem cell niches, in modulating cell fate. Thus, the ability to precisely tune and control extracellular cell/organelle shape and geometry in 2D and 3D, is of critical importance. PRIMO has been incredibly useful in this regard!

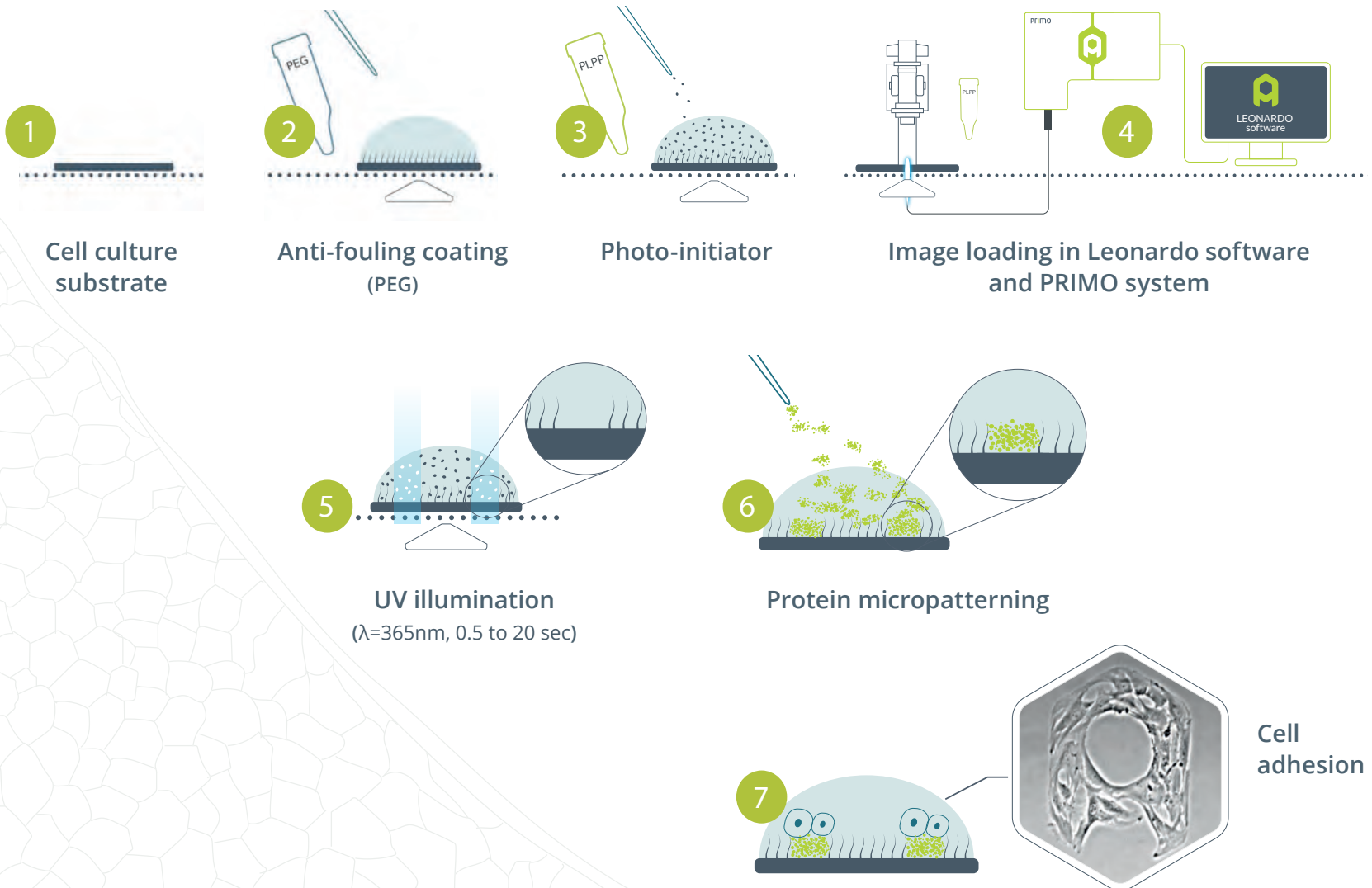
Yekaterina A. Miroshnikova

Human Frontiers Postdoctoral Fellow

University of Helsinki, Helsinki Institute of Life Science & Max Planck Institute for Biology of Ageing, Wickström Lab

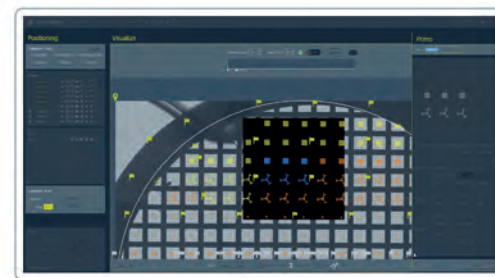
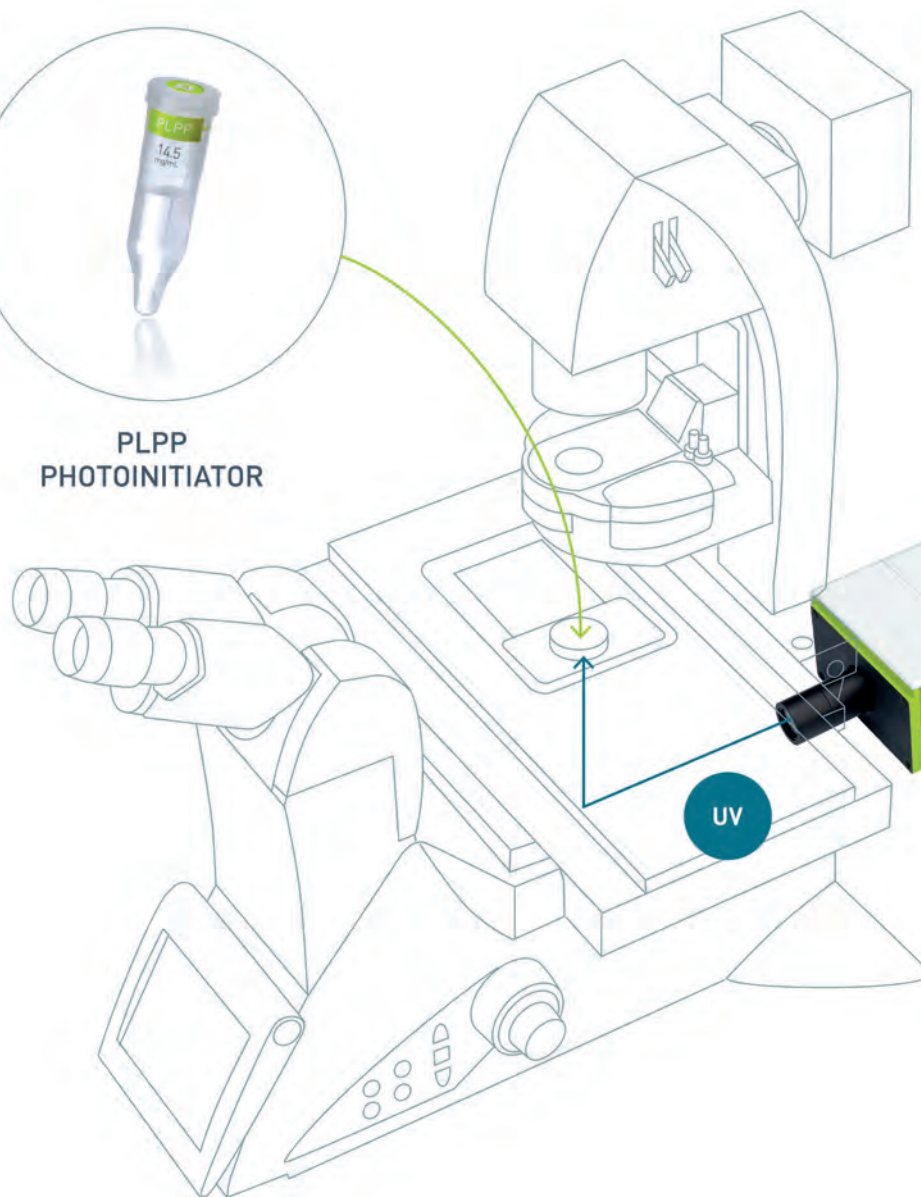


Micropatterning made easy





PLPP
PHOTOINITIATOR



LEONARDO SOFTWARE



PRIMO

Unrivalled performance

GRADIENTS

256

gray levels

MULTI-PROTEIN

3

depending on experimental conditions

Range of 10+ proteins used daily by our users

HIGH RESOLUTION

1.2 μm

over the entire illuminated field*

**Approximately 500x300 μm , 20x objective.*

ALIGNMENT

on microstructures*
or micropatterns

**Automatic detection
and patterns positioning*

COMPATIBLE

standard
substrates*

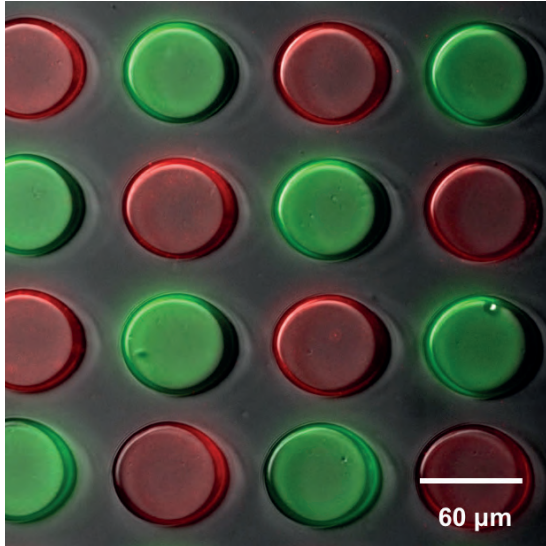
**stiff or soft, flat
or microstructured.*

FAST

0.5 sec

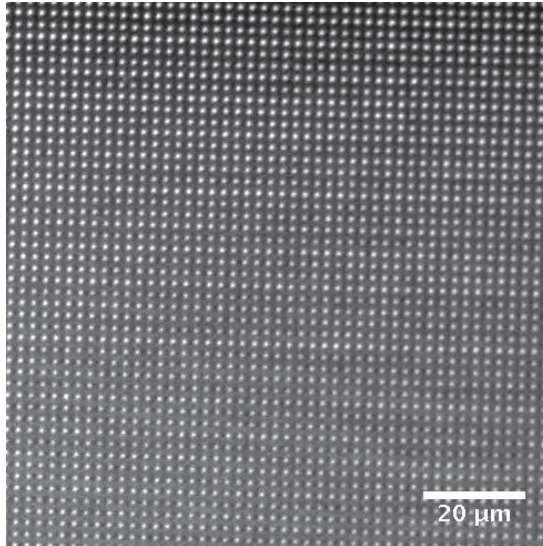
for a full field pattern*

**Approximately 500x300 μm ,
20x objective, with PLPP Gel.*



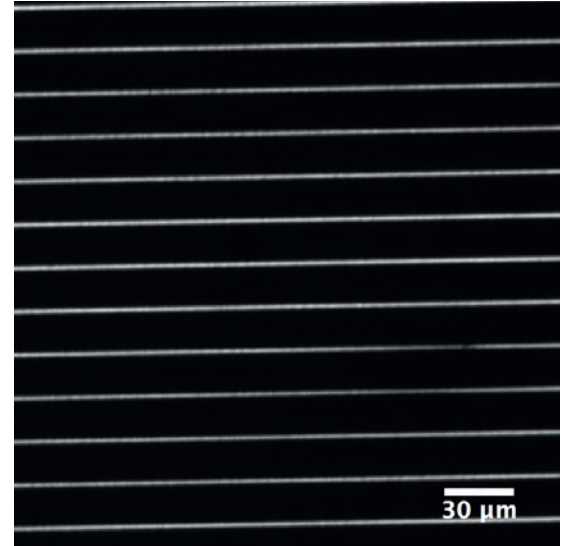
ALIGNMENT & MULTI-PROTEIN:

Sequential photopatterning of Fibrinogen-A488 in green and Protein A-A647 in red onto PDMS micropillars microfabricated with PRIMO.



HIGH RESOLUTION:

Epifluorescence microscopy image of 1,5μm dots (spaced by 1,5μm) of ProteinA-488 on PDMS.



HIGH RESOLUTION:

Epifluorescence microscopy image of 2 μm horizontal lines of ProteinA-488 on glass.

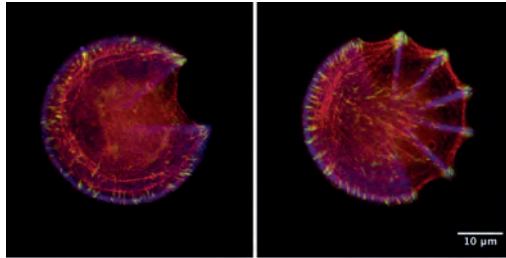


GRADIENTS:

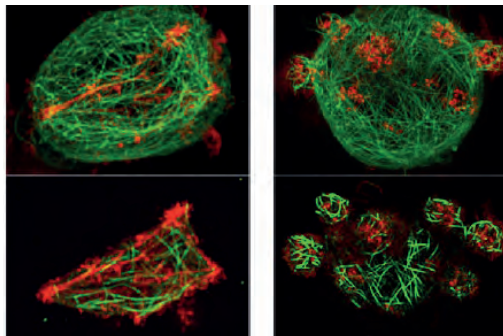
Epifluorescence microscopy image of a gradient of Fibrinogen-A488 on a glass coverslip.

Applications

Cytoskeleton studies

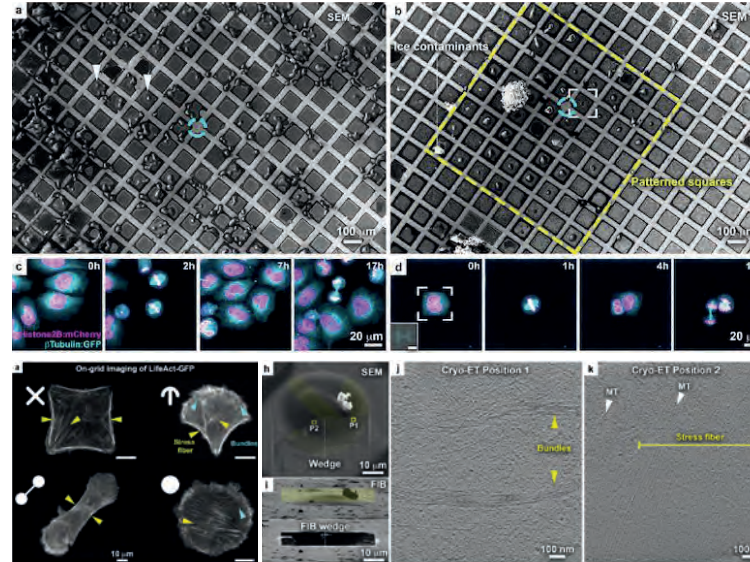


Embryonic fibroblasts from vimentin knockout mice on a fibronectin + fibrinogen pattern (blue), actin (red), focal adhesions (green). Courtesy of A.J. Jimenez and B. Vianay, Physics of cytoskeleton & Morphogenesis lab.



«A mechano-signalling network linking microtubules, myosin IIA filaments and integrin-based adhesions» N.B.M. Rafiq et al., Nat. Mat., 2019

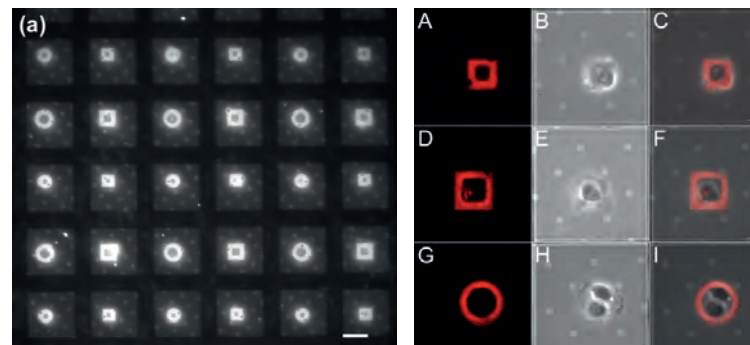
Cryo-ET cell sample preparation



(a) HeLa cells on standard gold-mesh grid SiO₂ holey film. Arrowheads: cells optimally positioned for FIB-milling. (b) HeLa cells on a gold-mesh grid with 20 µm diameter disk fibronectin patterns. (c-d) HeLa cells seeded on a (c) control and (d) patterned (H-shaped) gold-mesh grids with SiO₂ (R1/20) holey film. FOV: one single grid square.

(j-k) Tomographic slices of positions 1 and 2 indicated in (h). Actin bundles and internal stress fibers -indicated in (a)- are found in locations expected according to the actin map in a crossbow-shaped RPE1 cell.

M. Toro-Nahuelpan et al., Nature Methods, 2019

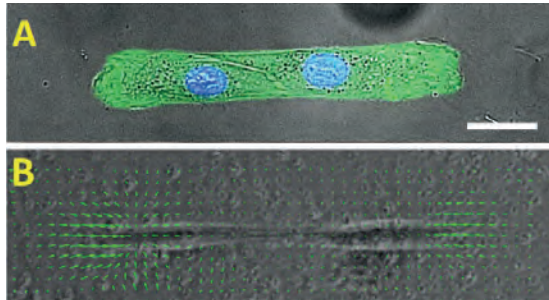


Left: Gelatin hollow squares and circles patterned between the gold grid bars of 200 mesh holey carbon EM grids. Scale bar, 50 µm.

Right: PtK1 cells plated on rhodamine-fibronectin square and patterns. Scale bar, 10 µm.

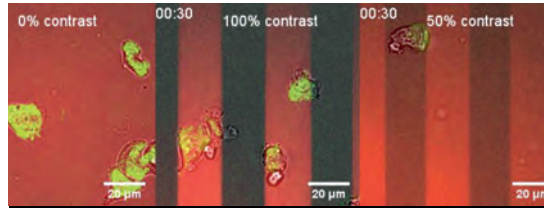
L. Engel et al., J. Micromech. Microeng., 2019

Force Measurement



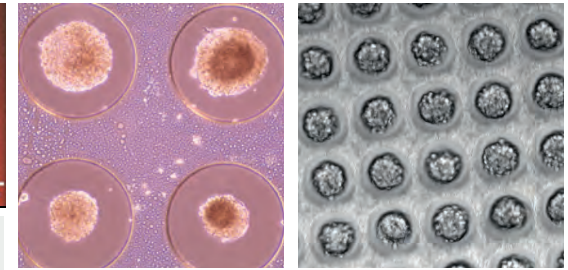
Two airway smooth muscle (ASM) cell ensemble on a rectangular gelatin micropattern (green) done with PRIMO on Nusil gel, scale bar = 25 μm .
S. R. Polio et al., Scientific Reports, 2019

Cell Migration



Left: negative control substrate with 0% adhesion contrast; Middle: positive control substrate with 100% adhesion contrast; Right: substrate with 50% adhesion contrast.
In green: cell adhesion patch shown by RICM; In red: substrate with higher adhesion; In grey: bright-field images showing cell body.
X. Luo et al., BioRxiv, 2019

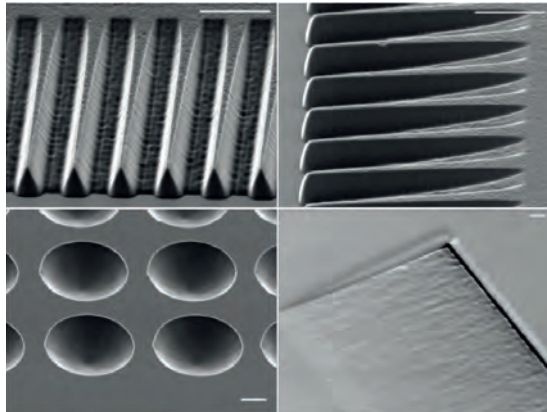
Spheroid Formation



Huh-7 cells forming spheroids on micropatterns of fibrinogen-A488 (wells $\text{Ø}=500 \mu\text{m}$, micropatterns $\text{Ø}=300 \mu\text{m}$).

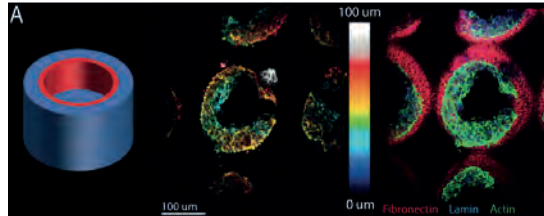
Spheroids of HEK cells in hydrogel microwells ($\text{Ø}=100 \mu\text{m}$, $H=175 \mu\text{m}$) photopolymerized with PRIMO. Courtesy of A. Pasturel and V. Studer.

Microfabrication



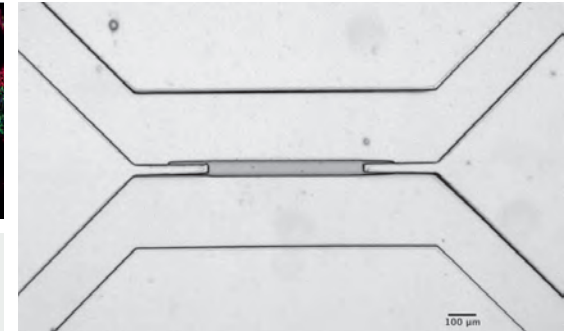
SEM images of structures microfabricated with ma-P 1275G resist and PRIMO. Scale bar = 30 μm

Hydrogel Structuration



Left panel: scheme of topographically (blue) and chemically (red) photopatterned hydrogels. Middle panel: COS-7 cells seeded on the gel (Z-scale). Right panel: patterned fibronectin (red), actin cytoskeleton (green) and nuclear envelope (blue).
A. Pasturel et al., BioRxiv, 2018.
doi.org/10.1101/370882

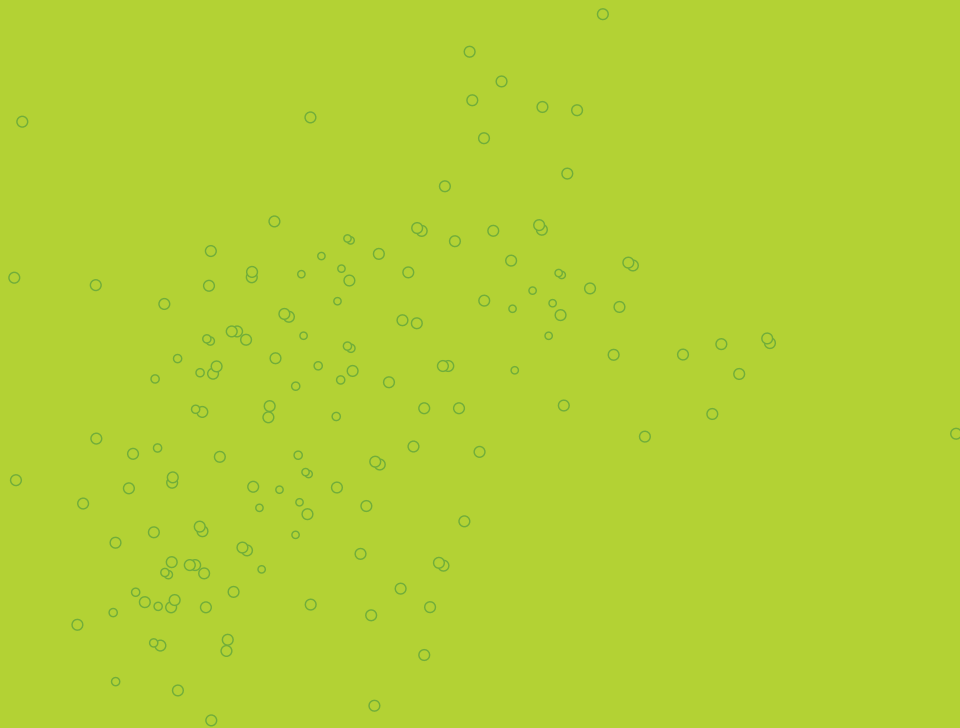
Microfluidics



Photopatterning with PRIMO system of pressure-resistant hydrogel-based permeable membrane within PEGDA microfluidic chips. Courtesy of J.-B. Salmon.

A complete bioengineering platform

We have developed complementary products to give you optimized and personalized control over your experimental conditions.





Nomos
Automation
of Experiments



Leonardo
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Software



PRIMO



Stencil
Multi-Well
Solution



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